# E-Nose System to Detect E-Coli in Drinking Water of Udupi District

Krishnamurthy Nayak<sup>1</sup>, Supreetha B.S.<sup>2</sup>, Deccaraman M.<sup>3</sup>, Vijayashree Nayak<sup>4</sup>,

<sup>1</sup>Research scholar, Dr. M.G.R. University, Chennai <sup>1.2</sup>Assistant Professor, Department of Electronics and Communication, MIT, Manipal <sup>3</sup>Dean, Department of Biotechnology, Dr. M. G. R. University, Chennai <sup>4</sup>Assistant Professor, Department of Biological Sciences, BITS Pilani, Goa

Abstract- E-nose (Electronic nose), which is an array of chemical sensors, is widely used in food industry and pollution control. Nowadays e-nose technology has become powerful tool to evaluate the aroma compounds during the quality control process. This work is intended to detect biological water contaminants like coliform group of bacteria in drinking water. However, a number of limitations are associated with traditional methods of microbial detection. The water samples were collected from the River Swarna in Udupi district and analyzed for the presence of E-coli. An E-nose system is developed using embedded PIC micro controller to detect and quantify the microbial contaminants. The results of this analysis were compared with standard Multiple Test Tube Fermentation (MTTF) method, which indicates the Most Probable Number (MPN) for the bacterial density in drinking water.

Keywords—E-nose, E-coli, PIC Micro Controller, Multiple Test Tube Fermentation (MTTF) method, Most Probable Number (MPN)

# INTRODUCTION

E-nose are being developed as systems for the automated detection and classification of odour, vapors and gases. The detection of gases using e-nose has a wide range of application in air quality monitoring [1], food spoilage detection [2] and medical diagnostic[3]. E-nose contains array of sensors resulting in multiple measurements which have been analyzed using pattern recognition techniques, An e-nose could be a universal vapor sensor , in practice however all existing e-nose detects only a family of related chemicals due to the difficulty of producing a variety of sensing materials required for universal sensing [4]. E-nose using SnO<sub>2</sub> gas sensors are used for detection of methanol contamination in whiskeys [5]. In this work, we detect E-coli in drinking water using  $CO_2$  sensor. The water is collected from Swarna river of Udupi district. Swarna river, which is born in Agumbe Ghats and flows through Udupi district, reaches Arabian sea. Swarna river water is the main source of drinking water for people residing in and around Manipal and Udupi.

Drinking water must possess high degree of purity and free from micro-organism. The potential and quality of drinking water is an economic resource and essential component for survival of living beings on earth. Living conditions are being deteriorated in major rural centers due to pollution caused by population explosion, urbanization and microbial contaminations [7]. The detection of microorganisms in drinking water that are harmful to health, gives a historical overview of the risk of disease from pathogens. E.coli is the most abundant coliform organism present in the human [8] and animal intestine, and can be present in faeces causing a main concern to the health. Since E.coli and other coliform organisms are present in large numbers in faeces and sewage, they can be detected in numbers as small as 1 in 100 ml of water. They are considered to be the most sensitive indicator bacteria for demonstrating excremental contamination. For this reason, quantification of E.coli in drinking water is important to assess the degree of pollution and thereafter to protect human health [6].

Traditionally indicator microorganisms have been used to suggest the presence of pathogens. The specific methods for the detection and enumeration of enteric pathogenic bacteria and viruses are complex and time consuming [11]. The test for detection and enumeration of indicator organisms, rather than pathogens are used.

### A. Assessment of Water Quality by Detecting Microbial Water Contaminants

I.

The commonly used methods to detect the presence of coliform bacteria are Membrane Filter technique and Multiple Test Tube Fermentation (MTTF) technique. Membrane filter technique involves direct plating for detection and estimation of coliform densities [7]. The toxic substances in the water may be absorbed by filter and interfaces with bacterial growth. The plate count provides an approximate enumeration of total number of bacteria multiplying at room temperature that provide approximate information about water quality. Thus, the membrane filter technique fails to detect and quantify bacterial count in drinking water. MTTF technique involves mixing different dilutions of a sample of water with lactose broth and incubating them in test tubes for 48 hours at room temperature. The presence of  $CO_2$  gas tubes will indicate the presence of coliform bacteria. Most Probable Number (MPN) represents the bacterial density [11]. The presence or absence test using MTTF method is used to monitor high quality samples, where the presence of the target organism is improbable. The sensitivity of this technique depends on the sample volume analyzed and the precision on the number of samples analyzed in parallel with each dilution step. In all of the enumeration techniques, the cultivation conditions are selected to

promote the multiplication of the target organisms, while simultaneously inhibiting the growth of other organisms. The balance between sensitivity and selectivity is the reason for different methods for drinking water and highly contaminated waters. Since in our work we are concentrating on drinking water, MTTF method is preferred. The MPN index represents the bacterial density.

## II. METHODOLOGY

Multiple Test Tube Fermentation (MTTF) method is performed with 100ml of water sample using three test tubes, containing serial dilution, that are incubated in to a broth media. The combinations of positive results are interpreted from statistical tables, to estimate the number of organisms present. After incubating water sample with media for 24 to 48 hours the MPN Index is calculated. If not more than 1 coliform is present per 100ml of water, then water is said to be safe for drinking. The experimental set up for MTTF test with e-nose system is shown in Fig.1.



Fig. 1 Experimental setup

The MTTF method, also referred as MPN method, is based on an indirect assessment of microbial density in the water sample by reference to statistical tables, to determine the MPN of organism present in the original sample. The MPN per 100ml of sample using three test-tubes of each dilution is recorded from the MPN (Table I). [APHA-AWWA-WPCF, 'Standard methods for the examination of water and wastewater']

The water samples collected for bacterial analysis are: Sample 1: Water collected directly from Swarna river during rainy season; Sample 2: Water collected directly from Swarna river during summer season; Sample 3: Water collected after preliminary filtration (Swarna river water); Sample 4: Filtered and chlorinated drinking water supplied from Udupi Municipal Corporation (Swarna river water).

The coliform group of bacteria is detected based on lactose fermentation. The release of  $CO_2$  gas at room temperature proportional to the bacterial count is the indication of bacterial contamination. To improve culture-based techniques for monitoring specificmicrobial populations in water samples, the current study has adapted the sensor technology [7] to automate the process of water analysis.

Number of positive tubes in dilutions			MPN per 100 ml	Numb positi dilutio	MPN per 100 ml		
10	1	0.1		10	1	0.1	
ml	ml	ml		ml	ml	ml	
0	0	0	0	2	0	0	9.1
0	1	0	3	2	0	1	14
0	0	2	6	2	0	2	20
0	1	0	3	2	0	3	26
0	1	1	6.1	2	1	0	15
0	1	2	9.2	2	1	1	20
0	1	3	12	2	1	2	27
0	2	0	6.2	2	1	3	34
0	2	1	9.3	2	2	0	21
0	2	2	12	2	2	1	28
0	2	3	16	2	2	2	35
0	3	0	9.4	2	2	3	42
0	3	1	13	2	3	0	29
0	3	2	16	2	3	1	36
0	3	3	19	2	3	2	44
1	0	0	3.6	3	0	0	23
1	0	1	7.2	3	0	1	39
1	0	3	11	3	0	2	64
1	1	0	7.3	3	0	3	95

# *TABLE I* MPN TABLE PER 100 ML OF SAMPLE

1	1	1	11	3	1	0	43
1	1	2	15	3	1	1	75
1	1	3	19	3	1	2	120
1	2	0	11	3	1	3	160
1	2	1	15	3	2	0	93
1	2	2	20	3	2	1	150
1	2	3	24	3	2	2	210
1	3	9	16	3	2	3	290
1	3	1	20	3	3	0	240
1	3	2	24	3	3	1	460
1	3	3	29	3	3	2	1100

The coliform group of bacteria is detected and quantified using PIC microcontroller based data acquisition system (DAQ). Fig. 2 shows the simplified schematic diagram of the e-nose system. The project flow is shown in Fig. 3.

Sampling system		Array of sensors		Computing <ul> <li>system</li> </ul>
--------------------	--	------------------	--	--------------------------------------

Fig.2. Simplified schematic diagram of e-nose system



Fig.3 Block diagram of e-nose system

The e-nose system involves hardware and software implementation. The water sample is tested using MTTF method and the  $CO_2$  gas released depending on the bacterial contamination, is sensed using array of nine carbon dioxide sensors. Signal conditioning circuit using OPAMP will improve the quality of signals. The sensor output voltage is measured using an operational amplifier with high input impedance and low bias current (less than 1 pA) shown in Fig. 4.





Fig.4 Sensor interface and signal conditioning

The PIC Microcontroller is programmed to detect and quantify the microbial water contaminants as shown in below flow diagram Fig. 5. The schematic diagram of e-nose system is built using PSPICE as shown in Fig. 6.







Fig.6. Circuit diagram of e-nose system

# EXPERIMENTAL RESULTS

III.

The MPN index of Bacterial count is recorded from statistical table and it is compared with sensor response of enose system as shown in Table II. The sensor response 220 mV for 350 ppm of  $CO_2$  gas is taken as threshold voltage. The tables prove that e-nose system accurately measures microbial water contaminants and indicates the degree of contamination. The bacterial concentration for all 4 samples are plotted as shown in Fig. 7, which indicates that bacterial count varies randomly and This system is able to monitor the degree of contamination and thus the water quality.

OF EACH DILUTION	TABLE II : TABLE OF BACTERIAL COUNT PER 100 ML OF SAMPLE USING THREE TEST-TUBES	3
	OF EACH DILUTION	_

MPN No (Index)					Sensor response				
Water sample	No of positive tubes out of 3			MPN	10ml	1ml	0.1ml	Percentag e Water	Comment
L	10 ml	1 ml	0. 1 ml	Index	$\begin{array}{c} {S_{11}} \\ {S_{12}} \\ {S_{13}} \end{array}$	$\begin{array}{c} {\bf S}_{21} \ {\bf S}_{22} \\ {\bf S}_{23} \end{array}$	$\begin{array}{c} {\bf S}_{31} \ {\bf S}_{32} \\ {\bf S}_{33} \end{array}$	Quality	
Sample1 (collected from river during rainy season)	3	3	2	1100/10 Oml	299m V 300m V 390m V	240m V 285m V 260m V	285m V 275m V 295m V	0%	Not Drinkable
Sample 2 (collected from river during summer)	2	2	2	35/ 100ml	293m V 328m V 215m V	289m V 300m V 205m V	229m V 329m V 219m V	25%	Not Drinkable
Sample 3(after preliminary filtration)	1	2	2	20 /100ml	282m V 210m V 200m V	280m V 260m V 199m V	500m V 290m V 203m V	50%	Treatment required
Sample 4(filtered and chlorinated by municipal corporation for drinking purpose)	0	0	0	0/100ml	120m V 118m V 110m V	115m V 116m V 115m V	105m V 108m V 110m V	100%	Drinkable

#### Sample1 -Not Drinkable





#### Sample 3 - Treatment Required

*Fig.7.* E-nose system responses for different water samples.

### IV. CONCLUSION

E-nose system found to be effective in detecting and accurately quantifying the microbial contaminants, specifically coliform group of bacteria (E. coli). This work compares the traditional bacterial count in MPN index with sensor response. The system monitors the quality of drinking water and display the percentage level of microbial contaminants. Further improvement in the system can be done by integrating advanced chemical sensors for detecting chemical contaminants and hardness of the drinking water.

# V. ACKNOWLEDGEMENT

The author wish to thank Manipal University for providing the research facilities at the environmental laboratory of MIT, Manipal.

### REFERENCES

- W. Bourgeoisis, A.C. Roman, J. Nicolas and R. M. Stutez, The use of sensor arrays for environmental monitoring: interests and limitation. Environmental Monitoring, 2003.
- [2]. T. Rajamakii, H. L. Alakomi, T. Ritvanen, E. Skytta, M. Smolander and R. Ahvenainen, Application of an E-nose for quality assessment of modified atmosphere packaged poultry meat, Food Control 17, 5-13, 2006..
- [3]. M. E. Shykhon, D.W. Morgan, R. Dutta, E.L. Hines and J.W. Gardner, Clinical evaluation of the electronic nose in the diagnosis of ear, nose and throat infection: a preliminary study. J. Laryngol. Otol. 118, 706-709, 2004.
- [4]. T. C. Pearce and S.M. Manuel, Chemical sensor array optimization: geometric and information theoretic approaches, in: T.C. Pearce, et al. (Eds.), Handbook of Machine Olfaction- E-nose technology, John Wiley & sons, 347-375, 2003.
- [5]. Chatchawal Wongchoosuk, Anurat Wisitisoraat, Adisorn Tauntranont and Teerakiat Kerdcharoen. "Portable electronic nose based on carbon nanotube-SnO<sub>2</sub> gas sensors and its application for detection of methanol contamination in whiskeys". Sensors and Actuators B: Chemical. Vol.147, pp.392-399,2010
- [6]. G. Craun, F. Nwachuku and N.Calderon, "Outbreaks in drinking water systems", Journal of Environmental Health. 1991-1998.
- [7]. Lindquist and P. Wide "Virtual water quality tests with an electronic tongue", Instrumentation and Measurement Technology Conference 2001, volume:ISBN:0-7803-68 2001.
- [8]. "Down to Earth, September", Science and Environment Fortnightly Centre for Science and Environment, New Delhi1-15,2008.
- [9]. Juan Rodriguez, Christian Duran and Adriana Reyes, "Electronic Nose for quality control of colomian coffee through the detection of defects in cup tests", Department of Electronic Engineering, University of Pamplona, Pamplona.2001.
- [10]. Rose. J. B "Water Quality Security", Environmental Science and Technology, 1995.
- [11]. Yehuda Kotti, "Estimation of Low numbers of Escherichia coli bacteriophage by use of Most Probable Number Method", Environmental Health Engg, University of Texas, Austin, 17 June 1965.
- [12]. Berg, G., "The Indicator Systems indicators of Bacteria in water and food" Ann Arbor Science Publishers, pp. 1–13, 1978.
- [13]. Kabler and S. I. Chang, "Quality aspects of water distribution systems", sanitary Eng, University of Illinosis, 1963.
- [14]. "A survey of the quality of water drawn from domestic wells in Nine Midwest States", CDC, September 1998.
- [15]. Syvang Xayyavong and Sandhya Babel, "Quality of Bottled Drinking Water", Case study in Vientiane ,Laos and Thaoland, International Conference of Life Sciences Institute.1978.
- [16]. Yang Jiejun and Wang Lin, "Assessment of surface and groundwater water quality at regional scale", A case study of Laoshan District, China, Environmental Science and Engg, Water Environmental Monitoring Centre of Qingdao, China, 2001.
- [17]. APHA-AWWA-WPCF, "Standard methods for the examination of water and wastewater", 3<sup>rd</sup> Edition, 1889.
- [18]. "A study in to the variation in microbiological water quality between different water source types" (MSc thesis), Cranfield University, School of Applied Sciences, 2008.
- [19]. Lindquist H. D., "Emerging pathogens of concern on drinking water", EPA publication, 1999.

# Biography



**K. M. Nayak** obtained his BE in Electronics and Communication Engineering from Mangalore University, Karnataka India and MTech in Industrial Biotechnology from Dr.MGR University Chennai. He is the Associate member of Institute of Engineers India and Life member of Indian society for Technical Education. Currently he is working as Assistant Professor (Sel. Grade) in Department of Electronics and Communication Engineering, MIT, Manipal of Manipal University and part time research scholar at Dr. MGR University, Maduravoyal, Chennai. His specialization includes Bio

MEMS and Microsensor and Bio Electronics. His current research includes E-nose technology for Environmental Monitoring and Artificial Neural Networks.